

presence of covalent hydrates for the ground- and excited-state chemistry of bipy and other aromatic N heterocycles and their coordination complexes cannot be ignored.

References and Notes

- (1) Research supported by National Science Foundation through Grant No. CHE76-21050.
- (2) (a) A. Jensen, F. Basolo, and H. M. Neuman, *J. Am. Chem. Soc.*, **80**, 2354 (1958); (b) R. H. Linnell and A. Kaczmarczyk, *J. Phys. Chem.*, **65**, 1196 (1961); (c) A. A. Schllt, *Anal. Chem.*, **35**, 1599 (1963).
- (3) R. D. Gillard, *Coord. Chem. Rev.*, **16**, 67 (1975).
- (4) M. Maestri, F. Bolletta, N. Serpone, L. Moggi, and V. Balzani, *Inorg. Chem.*, **15**, 2048 (1976).
- (5) C. Creutz and N. Sutin, *Proc. Natl. Acad. Sci.*, **72**, 2858 (1975).
- (6) G. Nord and O. Wernberg, *J. Chem. Soc., Dalton Trans.*, 866 (1972).
- (7) J. F. Endicott, Microsymposium on Photochemistry and Photophysics of Coordination Compounds, Ferrara, Italy, July 1976.
- (8) J. Jousset-Dubien and J. Houdard-Pereyre, *Bull. Soc. Chim. Fr.*, 2619 (1969); K. E. Wilzbach and D. J. Rausch, *J. Am. Chem. Soc.*, **94**, 2178 (1970).
- (9) A. Albert, *Adv. Heterocycl. Chem.*, **20**, 117 (1976).
- (10) F. H. Estheimer and O. T. Benfey, *J. Am. Chem. Soc.*, **78**, 5309 (1956); K. Nakamoto, *J. Phys. Chem.*, **64**, 1420 (1960); W. A. E. McBryde, *Can. J. Chem.*, **43**, 3472 (1965).
- (11) (a) Y. Gondo, *J. Chem. Phys.*, **41**, 3928 (1964); (b) G. M. Badger and I. S. Walker, *J. Chem. Soc.*, 122 (1956); (c) L. Gil, E. Moraga, and S. Bunel, *Mol. Phys.*, **12**, 333 (1967).
- (12) T. M. Spotswood and C. I. Tanzer, *Aust. J. Chem.*, **20**, 1227 (1967).
- (13) The difference in the absorbance of the 281-nm band is the result of the dependence of ϵ upon solvent medium.¹⁴
- (14) I. B. Berlman, "Handbook of Fluorescence Spectra of Aromatic Molecules", Academic Press, New York, N.Y., 1971.
- (15) M. Kasha, *Discuss. Faraday Soc.*, **9**, 14 (1950).

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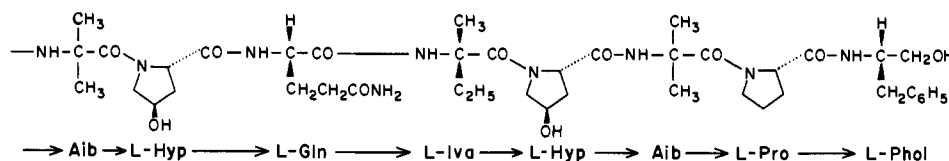
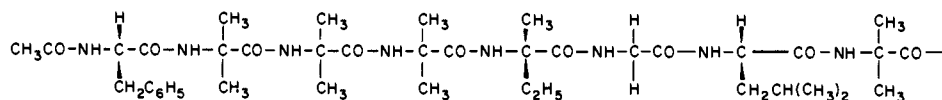
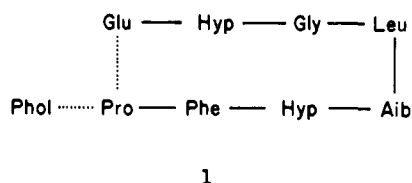
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Structure of Antiamoebin I from High Resolution Field Desorption and Gas Chromatographic Mass Spectrometry Studies

Sir:

The antibiotic antiamoebin was reported in 1968^{2a} to be produced by *Emericellopsis poonensis* Thirum., *E. synnematicola* Mathur and Thirum., and *Cephalosporium pimprina* Thirum., and to be active against protozoa and helminths.^{2b} Subsequently, its structure was assigned as **1**, a

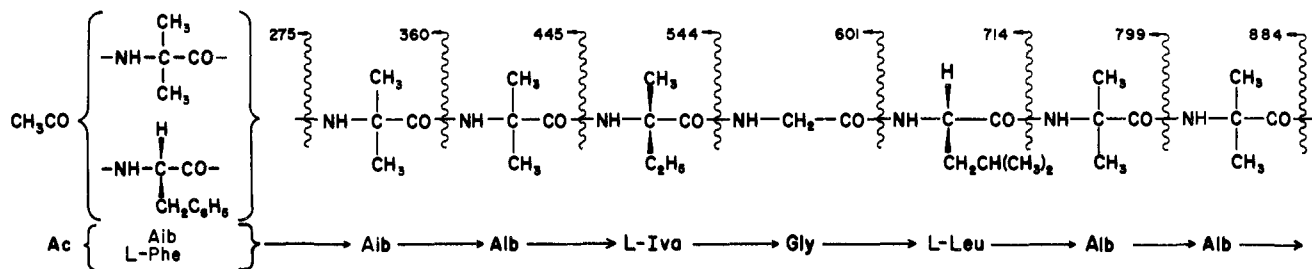


cyclic octapeptide linked to phenylalaninol.³ Oleic, linoleic, and palmitic acids were also indicated to be a part of the molecule.⁴ We have now shown that antiamoebin is a mixture of two closely related compounds, antiamoebins I (~98%) and II (~2%), separable by countercurrent distribution in the system $\text{CHCl}_3\text{-C}_6\text{H}_6\text{-MeOH-H}_2\text{O}$, 30:30:46:14,³ in which we find $K_I = 5.7$ and $K_{II} = 2.1$, and that the structures of these antibiotics are actually quite different from **1**.

We assign here structure **2** to antiamoebin I, employing gas chromatography-high resolution electron impact mass spectrometry (GC-HREIMS) and high resolution field desorption mass spectrometry (HRFDMS) as principal structural tools. We also propose the name peptaibophol (defined as peptide antibiotics containing phenylalaninol and several moles of α -aminoisobutyric acid as well as other amino acids) for this class of antibiotics, which we have recently shown includes emerimicins II, III, and IV,⁵ zervamicins I and II,^{5b} and alamethicins I and II,⁶ as well as antiamoebins I and II. Two other antibiotics, suzukacillin⁷ and samarosporin,⁸ also belong to this class, as does (probably) stilbellin.⁹

Antiamoebin I, mp 194–196 °C (MeOH-H₂O), $[\alpha]^{25}_D +17.8^\circ$ (*c* 2.1, MeOH), gives microanalyses agreeing with the molecular formula $\text{C}_{82}\text{H}_{127}\text{N}_{17}\text{O}_{20}\cdot 2\text{H}_2\text{O}$ (mol wt (anhydrous), 1669), while the cationated molecular ion¹⁰ is found at *m/e* 1692 (*M* + Na) in the field desorption mass spectrum (FDMS).¹¹ Acetylation of antiamoebin I with acetic anhydride-pyridine gave antiamoebin I triacetate, whose FDMS showed ions at *m/e* 1818.9596 (matched vs. the peak at *m/e* 1720.9514 ($\text{C}_{34}\text{H}_{18}\text{F}_{56}\text{N}_3\text{O}_6\text{P}_3$) in the FDMS of bis(dodecafluoroheptoxy)tetrakis(octafluoropentoxy)cyclotriphosphazene),^{11c} 1758, and 1698, corresponding to $\text{C}_{88}\text{H}_{133}\text{N}_{17}\text{O}_{23}\text{Na}$ (*M* + Na), *M* + Na – HOAc, and *M* + Na – 2HOAc, respectively. Cationated molecular ions (*M* + Na) for the tripropionate and tributryate were found by FDMS at 1860 and 1902, respectively, while those for the lithium and potassium conjugates of the triacetate were found at *m/e* 1802 and 1834, respectively.¹⁰

Hydrolysis of antiamoebin I with 6 N hydrochloric acid gave a mixture of amino acids but no fatty acids. Analysis of the hydrolysis mixture was carried out by FDMS followed by HRFDMS (employing a variety of reference standards described elsewhere),⁶ which gave *M* + H ions at *m/e* 76.0409 ($\text{C}_2\text{H}_6\text{NO}_2$, Gly), 104.0714 ($\text{C}_4\text{H}_{10}\text{NO}_2$, Aib, α -aminoisobutyric acid;^{12,13} cf. seq.), 116.0721 ($\text{C}_5\text{H}_{10}\text{NO}_2$, Pro), 118.0883 ($\text{C}_5\text{H}_{12}\text{NO}_2$, Iva, isovaline, α -amino- α -methylbutyric acid;^{14,15} cf. seq.), 132.0654 ($\text{C}_5\text{H}_{10}\text{NO}_3$, Hyp), 132.1012 ($\text{C}_6\text{H}_{14}\text{NO}_2$, Leu), 148.0604 ($\text{C}_5\text{H}_{10}\text{NO}_4$, Glu), 152.1078 ($\text{C}_9\text{H}_{14}\text{NO}$, Phol, phenylalaninol; cf. seq.), and 166.0849 ($\text{C}_9\text{H}_{12}\text{NO}_2$, Phe); by amino acid analysis (Beckman/Spinco, Model 120), which indicated 6–7 mol of Aib, 2

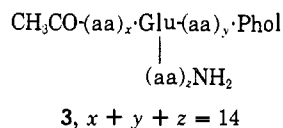


a

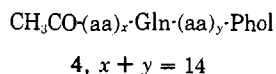
mol of Hyp, and 1 mol each of Gly, Leu, Pro, Phe, Glu, and Phol;^{10a} and by GC-HREIMS of the *n*-butyl and methyl ester derivatives of the *N*-trifluoroacetyl amino acids, which indicated 5–7 mol of Aib, 2 mol each of Iva and Hyp, and 1 mol each of Gly, Leu, Phol, Pro, Phe, and Glu (all identified by comparison of retention times and fragmentation patterns with those of authentic samples). Combining these results indicates the composition 5–7 Aib, 2 Iva, 2 Hyp, and 1 each of Gly, Leu, Phol, Pro, Phe, and Glu.

Analysis of the *N*-trifluoroacetyl amino acid methyl esters on a column of 10% *N*-lauroyl-*N'*-*tert*-butyl-L-valinamide on 60–80 mesh Chrom WAW,^{16,17} employing somewhat modified conditions,⁵ indicated that all of the optically active amino acids have the L configuration. Phol was isolated and also found to have the L configuration:¹⁸ $[\alpha]^{23}_D -23.5^\circ$ (*c* 2.47, MeOH) (lit. $[\alpha]^{24}_D -23.3^\circ$ (*c* 2.01, MeOH)).¹⁹

Antiamoebin I does not react with reagents for amino (N terminal) or carboxyl (C terminal) groups. Thus, either it is a cyclic peptide in which there are no N-terminal or C-terminal amino acids (and the γ -carboxyl group of Glu and the amino group of Phol are present in amide linkages) or it is a linear peptide in which the N-terminal and C-terminal groups occur derivatized. Antiamoebin I analyzes for one $-\text{CONH}_2$ group on hydrolysis to ammonia, and its ¹H NMR spectrum shows a singlet at 1.91 ppm (3 H, s) assignable to an acetyl methyl group. The presence of acetyl, primary amide, and three esterifiable hydroxyl groups and the mol wt 1669 argue that antiamoebin I is a linear peptide, as in 3, composed of 1 mol each of Gly, Leu, Pro, Glu, Phe, and Phol, 2 each of Iva and Hyp, and 6 of Aib. In agreement with this structure, 17 amide carbonyl carbons can be observed in the ¹³C NMR spectrum (DMSO-*d*₆) of antiamoebin I: at 175.9, 175.7, 175.6, 175.2, 174.9, 173.5, 173.2, 173.0, 172.9, 172.5, 172.2, 171.8 (2), 171.7, 170.8, and 170.5 (2) ppm.

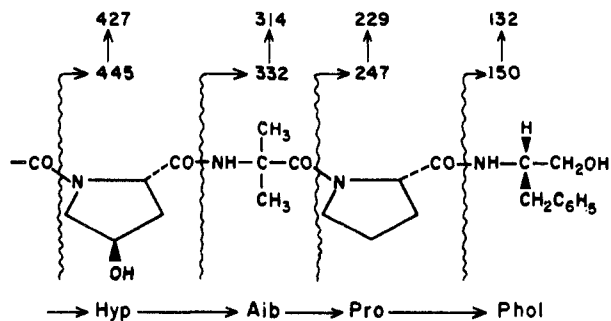


Dehydration of antiamoebin I with ethylene chlorophosphite in triethyl phosphite followed by reduction (Na-NH₃-MeOH),²⁰ hydrolysis (6 N HCl, 110 °C, 24 h), derivatization (*N*-trifluoroacetyl *n*-butyl ester), and analysis of the derivatives by GC and GC-EIMS did not show any glutamic acid but showed the presence of ornithine in the mixture. This clearly indicated that Glu is present as Gln in antiamoebin I. Structure 3 can then be written as 4.



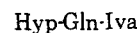
The order of linkage of the amino acids was established by mass spectrometry. The HREIMS of antiamoebin I shows characteristic CO-N cleavage peaks (the major peaks in the spectrum) at *m/e* 884.5327 (C₄₄H₇₀N₉O₁₀), 799.4770

(C₄₀H₆₃N₈O₉), 714.4209 (C₃₆H₅₆N₇O₈), 601.3374 (C₃₀H₄₅N₆O₇), 544.3163 (C₂₈H₄₂N₅O₆), 445.2471 (C₂₃H₃₃N₄O₅), 360.1968 (C₁₉H₂₆N₃O₄), and 275.1217 (C₁₅H₁₉N₂O₃) for the N-terminal sequence, and a weak CO-N cleavage peak at 190.0850 (C₁₁H₁₂N₂O₂), thus establishing the partial structure a. Similar, generally smaller peaks at 445.2453 (C₂₃H₃₃N₄O₅), 427.2374 (C₂₃H₃₁N₄O₄), 332.1951 (C₁₈H₂₆N₃O₃), 314.1893 (C₁₈H₂₄N₃O₂), 247.1431 (C₁₄H₁₉N₂O₂), 229.1341 (C₁₄H₁₇N₂O), 150.0915 (C₉H₁₂NO), and 132.0814 (C₉H₁₀N) establish the C-terminal sequence and the partial structure b. The peaks at *m/e* 884 and 799 are also present in the FDMS of antiamoebin I and its acetate.



b

Additional and confirmatory peptide sequences come from partial hydrolysis of antiamoebin I: (i) in 12 N hydrochloric acid-acetic acid (1:1), 25 °C, 72 h; (ii) in 6 N hydrochloric acid in methanol, 70 °C, 6 h. Each mixture of oligopeptides formed was analyzed in two ways. First, it was converted to the corresponding mixture of *N*-trifluoroacetyl *n*-butyl (or methyl) ester derivatives and analyzed by GC-HREIMS and GC-FIMS (FI, field ionization); peptides identified from their molecular formulas²¹ and fragmentation patterns in this way are marked with an asterisk (*). Second, the mixture of peptides was analyzed by HRFDMS;²² peptides identified from their protonated molecular ions in this way are marked with a dagger (†). A partial list of the peptides found (Ac-Phe-Aib-Aib-Aib-Iva† (562.3234, C₂₈H₄₄N₅O₇), Ac-Phe-Aib-Aib-Aib† (463.2499, C₂₃H₃₅N₄O₆), Phe-Aib*, Phe-Aib-Aib*, Aib-Aib*, Aib-Iva*, Iva-Gly-Leu*, Gly-Leu-Aib-Aib† (359.2283, C₁₆H₃₁N₄O₅), Gly-Leu*, Gly-Leu-Aib*, Leu-Aib*, Leu-Aib-Aib*† (302.2054, C₁₄H₂₈N₃O₄), Hyp-Gln-Iva† (359.1921, C₁₅H₂₇N₄O₆), Glu(α)-Iva*, Hyp-Aib-Pro*, Pro-Phol*† (249.1625, C₁₄H₂₁N₂O₂)) confirms a and b and establishes c. Thus, only one structure for antiamoebin I is allowed: a-c-b (2).²²



c

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provided by the National Science Foundation. We are grateful to Dr. M. J. Thirumalachar, former Superintendent of Research, Hindustan Antibiotics, Ltd., for the sample of anti-amoebin.

References and Notes

- (1) Presented in part at the 9th International Symposium on the Chemistry of Natural Products, IUPAC, Ottawa, Canada, June 24–28, 1974, Abstract 1B.
- (2) (a) M. J. Thirumalachar, *Hindustan Antibiot. Bull.*, **10**, 287–289 (1968). (b) Anti-amoebin has very recently been shown (P. Mueller, personal communication to K. L. Rinehart, Jr.) to have alamethicin-like activity in altering membrane permeability.
- (3) M. G. Vaidya, P. V. Deshmukh, and S. N. Chari, *Hindustan Antibiot. Bull.*, **11**, 81–89 (1968).
- (4) P. V. Deshmukh, *Hindustan Antibiot. Bull.*, **10**, 299–302 (1968).
- (5) (a) R. C. Pandey, J. C. Cook, Jr., and K. L. Rinehart, Jr., *J. Am. Chem. Soc.*, following paper in this issue; (b) R. C. Pandey and K. L. Rinehart, Jr., unpublished results.
- (6) R. C. Pandey, J. C. Cook, Jr., and K. L. Rinehart, Jr., *J. Am. Chem. Soc.*, in press.
- (7) G. Jung, W. A. König, D. Leibfritz, T. Ooka, K. Janko, and G. Boehm, *Biochim. Biophys. Acta*, **433**, 164–181 (1976).
- (8) N. Inoue, A. Inoue, M. Furukawa, and N. Kanda, *J. Antibiot.*, **29**, 618–622 (1976).
- (9) (a) Alb, Iva, and Phol are not detected under the usual conditions. (b) Thus, the ratio of Hyp (3) to Gly, Leu, Pro, Phe, and Glu (2 each) reported for stilbellin by amino acid analysis (K. Sasaki, H. Minato, K. Katagiri, S. Hayakawa, and T. Matsushima, *J. Antibiot.*, **24**, 67–68 (1971)) suggests that stilbellin might be identical with anti-amoebin.
- (10) K. L. Rinehart, Jr., J. C. Cook, Jr., H. Meng, K. L. Olson, and R. C. Pandey, *Nature*, in press.
- (11) (a) H. D. Beckey in "Biomedical Applications of Mass Spectrometry", G. R. Waller, Ed., Wiley-Interscience, New York, N.Y., 1972, pp 795–816; (b) K. L. Rinehart, Jr., J. C. Cook, Jr., K. L. Maurer, and U. Rapp, *J. Antibiot.*, **27**, 1–13 (1974); (c) K. L. Olson, K. L. Rinehart, Jr., and J. C. Cook, Jr., *Biomed. Mass Spectrom.*, **4**, in press.
- (12) G. W. Kenner and R. C. Sheppard, *Nature*, **181**, 48 (1958).
- (13) C. E. Meyer and F. Reusser, *Experientia*, **23**, 85–86 (1967).
- (14) F. Ehrlich and A. Wendel, *Biochem. Z.*, **8**, 438–466 (1908).
- (15) C. G. Baker, S. C. J. Fu, S. M. Birnbaum, H. A. Sober, and J. P. Greenstein, *J. Am. Chem. Soc.*, **74**, 4701–4702 (1952).
- (16) E. Gil-Av and B. Feilbush, *Tetrahedron Lett.*, 3345–3347 (1967).
- (17) R. Charles, U. Beittler, B. Feilbush, and E. Gil-Av, *J. Chromatog.*, **112**, 121–133 (1975).
- (18) P. V. Deshmukh and M. G. Vaidya, *Nature*, **217**, 849 (1968).
- (19) R. S. Kapil, B. C. Gautam, M. M. Vohra, and N. Anand, *Indian J. Chem.*, **4**, 177–187 (1966).
- (20) C. Ressler and D. V. Keshelkar, *J. Am. Chem. Soc.*, **88**, 2025–2035 (1966).
- (21) Molecular formulas determined by HRFDMS suggest the amino acid compositions for the oligopeptides, though they cannot, of course, sequence the amino acids in the peptides. The sequences shown for the peptides marked with a dagger are obtained by combining the HRFD data with the partial structures a and b, with the exception of the tripeptide Hyp-Gln-Iva, whose sequence was assigned from GC-MS data. That key tripeptide has also been isolated under the same hydrolysis conditions (II) and characterized by IR and ¹H NMR spectra, and its sequence has been established by direct probe HREIMS.
- (22) Although no peptides were observed containing Alb-Hyp and Iva-Hyp linkages, the molecular formula of anti-amoebin I allows no additional amino acids and partial structures a, b, and c account for the entire molecule.

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Structures of the Peptide Antibiotics Emerimicins III and IV^{1,2}

Sir:

The emerimicins, produced by *Emericellopsis microspora* in the presence of *trans*-4-*n*-propyl-L-proline (propylproline), are peptide antibiotics with moderate activity against gram-positive bacteria and against protozoa.³ In the present report we assign structures **1** to emerimicin IV, the principal com-

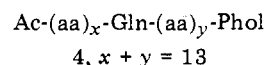
ponent, and **2** to emerimicin III, employing mainly field desorption mass spectrometry (FDMS), gas chromatography-high resolution electron impact mass spectrometry (GC-HREIMS) and gas chromatography-field ionization mass spectrometry (GC-FIMS). We also relate the structures of these antibiotics to that of anti-amoebin I (**3**),^{1b} another member of this class of antibiotics for which we propose the name peptaibophols.⁴

Emerimicin IV (mp >200 °C dec, pure by HPLC) has mol wt 1572, established by its field desorption mass spectrum (FDMS)⁵ after cation exchange,⁶ which shows M + Na and M + K ions at *m/e* 1595 and 1611, respectively. This was confirmed by the FDMS of cationated emerimicin IV triacetate (obtained from emerimicin IV by acetylation with acetic anhydride-pyridine at room temperature and purification on silica gel), which shows ions at *m/e* 1721 (M + Na), 1661 (M + Na – HOAc), and 1601 (M + Na – 2HOAc), and at 1737 (M + K), 1677 (M + K – HOAc), and 1617 (M + K – 2HOAc).

On vigorous acidic hydrolysis (6 N aqueous HCl, 110 ± 1 °C, 24 h) emerimicin IV gave glycine (Gly), α-aminoisobutyric acid (Aib), valine (Val) and/or isovaline (Iva), hydroxyproline (Hyp) and/or leucine (Leu), glutamic acid (Glu), phenylalanine (Phol), and phenylalanine (Phe), all identified from their molecular ions in an FDMS^{1b} of the hydrolysate. Quantitation of the amino acids on an amino acid analyzer (Beckman/Spinco, Model 120) indicated the molar ratio 7.0 ± 0.3 Aib:1 Gly:1 Val:1 Leu:1 Phol:2 Hyp:1 Phe:1 Glu, while gas chromatographic and GC-EIMS analysis^{1b} of the *N*-trifluoroacetyl derivatives of the *n*-butyl esters indicated the molar ratio 6.5 ± 1.3 Aib:1 Iva:1 Gly:1 Val:1 Leu:1 Phol:2 Hyp:1 Phe:1 Glu.

Emerimicin IV contains no carboxyl, carbalkoxyl, or primary amino group but analyzes for one –CONH₂ group per mole, while the ¹H NMR spectrum shows an acetyl (acetamido) methyl group at 1.88 ppm.^{1b} With an acetyl group, a terminal amide, 1 mol each of Gly, Val, Iva, Leu, Phe, Glu, and Phol, 2 mol of Hyp, and 6–8 mol of Aib, the mol wt 1572 indicates that emerimicin IV has the molecular formula C₇₇H₁₂₀N₁₆O₁₉ and contains 6 mol of Aib. Microanalyses reported earlier³ agree with this formula, as the dihydrate, while the ¹³C NMR spectrum of emerimicin IV contains the expected 16 carbonyl carbons, located at 175.9, 175.7, 175.0, 174.9, 173.8, 173.7, 173.2, 172.4, 172.2, 172.1, 171.9, 171.7, 171.5, 170.3 (2), and 170.0 ppm.

Emerimicin IV, like anti-amoebin I,^{1b,8} on dehydration (ethylene chlorophosphate/triethyl phosphite, 100 ± 1 °C, 24 h), reduction (Na–NH₃–MeOH), and hydrolysis (6 N HCl, 110 ± 1 °C, 24 h) followed by derivatization (*N*-trifluoroacetyl *n*-butyl esters) did not give any Glu but gave a new amino acid identified as ornithine (Orn) by GC and GC-MS. Thus, the primary amide group is present in Gln in emerimicin IV and the general structure **4** can be written.



The HREIMS of emerimicin IV and its triacetate contain peaks (all intense save that at *m/e* 190) for the sequence a at 190.0865 (C₁₁H₁₂NO₂), 275.1396 (C₁₅H₁₉N₂O₃), 360.1926 (C₁₉H₂₆N₃O₄), 445.2448 (C₂₃H₃₃N₄O₅), 544.3133 (C₂₈H₄₂N₅O₆), 601.3344 (C₃₀H₄₅N₆O₇), 714.4177 (C₃₆H₅₆N₇O₈), 799.4682 (C₄₀H₆₃N₈O₉), and 884.5242

1 (emerimicin IV): Ac-L-Phe-Aib-Aib-Aib-L-Val-Gly-L-Leu-Aib-Aib-L-Hyp-L-Gln-L-Iva-L-Hyp-Aib-L-Phol

2 (emerimicin III): Ac-L-Phe-Aib-Aib-Aib-L-Val-Gly-L-Leu-Aib-Aib-L-Hyp-L-Gln-L-Iva-L-Hyp-L-Ala-L-Phol

3 (anti-amoebin I): Ac-L-Phe-Aib-Aib-Aib-L-Iva-Gly-L-Leu-Aib-Aib-L-Hyp-L-Gln-L-Iva-L-Hyp-Aib-L-Pro-L-Phol